

In situ hybridization on EDC-fixed FFPE tissues using DIG-labeled miRCURY LNA™ detection probes

By Dr. Liming Luan, the Thomas Andl Laboratory, Vanderbilt University, Nashville, USA.

Deparaffinize the sections

1. Xylene, 3 X 5 min.
2. Ethanol, 100% 2 X 5 min.
3. Ethanol, 70% 5 min.
4. Ethanol, 50% 5 min.
5. Ethanol, 25% 5 min.
6. RNase-free H₂O, 1 min.

Deproteinate the sections

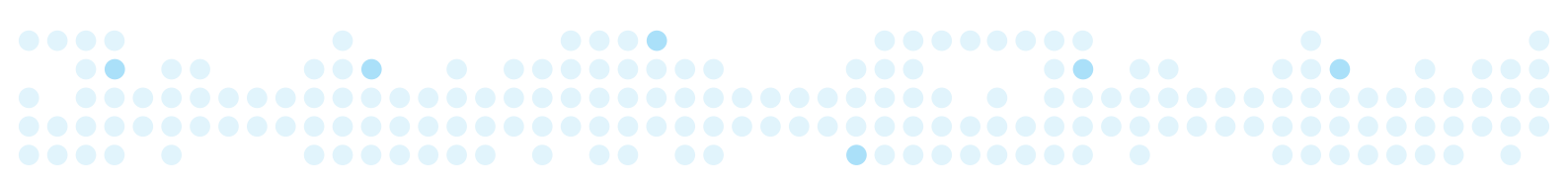
7. Wash slides with PBS, 2 X 5 min.
8. Soak slides in 75 ml PBS containing 20 µg/ml Prot. K (P4850, Sigma), 20 min.
9. Wash slides with PBS, 2 X 5 min.
10. Fix sections with fresh 4% PFA (in PBS), 10 min.
11. Wash slides with 0.2% Glycine (in PBS), 5 min.
12. Wash slides with PBS, 2 X 5 min.

EDC fixation

13. Incubate slides in 75 ml of freshly prepared imidazole buffer, 2 X 10 min.
14. Circle the sections with a DAKO pen.
15. Place slides in a humidified chamber (dH₂O), add 500 µl freshly prepared EDC solution to each slide and incubate for 1-2 h at room temperature.
16. Wash slides with 0.2% Glycine (in PBS), 5 min.
17. Wash slides with PBS, 2 X 5 min.

Prehybridization and hybridization

18. Add 500 µl prehybridization buffer (50% Formamide, 5X SSC) to each slide, room temperature, 15 min.
19. Denature the probe at 85°C for 3 min, chill on ice.
20. Add probe (final concentration, 20 nM) in hybridization buffer [50% Formamide, 5X SSC, 10% Dextran Sulphate Sodium (DSS), 500 µg tRNA, 0.02% BSA].
21. Add 200 µl hybridization mix per slide and cover with baked cover glasses.
22. Hybridize sections in a humidified chamber (50% Formamide, 5X SSC) for 16-18 hours.
Hybridization temperature = DNA T_m probe - 21°C.



Stringency Wash

23. Wash slides in 5X SSC, 50% Formamide at hybridization temperature, 15 min.
24. Wash slides in 2X SSC, 50% Formamide at hybridization temperature, 30 min.
25. Wash slides in 0.2X SSC, 50% Formamide at hybridization temperature, 2 x 30 min.
26. Rinse slides in 0.2X SSC at room temperature, 5 min.

Immunological Detection

27. Soak slides in Buffer I [150 mM NaCl, 100 mM Tris-Cl (pH7.5)] at room temperature for 5 min.
28. Add 200 µl of blocking buffer [1% Blocking reagent (Roche) in Buffer I] to each slide, room temperature, 1 hr.
29. Antibody incubation [anti-DIG-AP Fab fragment (Roche), 1:5000, in blocking buffer], 4°C, overnight.
30. Wash slides in Buffer I, 3 x 5 min.
31. Wash slides in Buffer II [100 mM NaCl, 100 mM Tris-Cl (pH9.5)], 5 min.

Color Development

32. Mix 200 µl of the NBT/BCIP Stock Solution (Roche) with 10 ml Buffer II.
33. Add diluted NBT/BCIP Solution to the slides and keep them in a light-protected humidified box, room temperature, 1-4 hr. For slow color development, transfer the box to 4°C, overnight.
34. Wash slides with PBS, 3 x 5 min.
35. Wash slides with dH₂O, 5 min.
36. Counter stain sections with Methyl green.

Reagents and Buffers

Imidazole buffer (160 ml, 0.13 M 1-methylimidazole, 300 mM NaCl, pH 8.0)

Add 1.6 ml of 1-methylimidazole to 130 ml water, adjust the pH by adding approximately 450 µl 12 M HCl to pH 8.0, then add 16 ml 3 M NaCl and water to a final volume of 160 ml.

EDC solution (0.16 M L-ethyl-3- (3-dimethylaminopropyl) carbodiimide (EDC))

Add 176 µl EDC into 10 ml of imidazole buffer, and then readjust the pH of the EDC solution by adding about 100 µl 12 M HCl to pH 8.0.

References

1. <http://www.exiqon.com/ls/Documents/Scientific/FFPE%20in%20situ%20hybridization.pdf>
2. Molecular cloning : a laboratory manual / Sambrook, Joseph; Russell, David W. 3rd ed. New York: CSHL, 2001.
3. Pena *et al.* miRNA *in situ* hybridization in formaldehyde and EDC-fixed tissues. *Nat. Methods.* 2009, 6: 139-41.

DIG: DIG is licensed from Roche Diagnostics GmbH.

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